Method 1

Appendix of Real-time quantitative PCR training course.

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This document explains how you can interpret the Ct values from $\Delta\Delta$ Ct method and how you can determine the $\Delta\Delta$ Ct method is applicable for your Experimental instead of the conventional calibration curve method.

If you are not sure which method to use, use calibration curve method because it is more reliable for most of the real-time PCR applications.

Experimentalal condition

- Sample subject: Control (Cont) and Experimental (Exp)
- Target gene: Tar
- Reference gene: *Ref*

Results

- Control sample Ct for Tar (ContCt Tar) was 22, Ct for Ref (ContCt Ref) was 19.
- Experimental sample Ct for Tar (ExpCt Tar) was 25, Ct for Ref (ExpCt Ref) was 18.

$$\Delta \text{Ct}^{Tar} = (2^{\text{ExpCt}^{Tar}} / 2^{\text{ContCt}^{Tar}})$$

= $(2^{25} / 2^{22}) = (2^{25-22}) = 2^{3}$

This calculation assumes that the Tar DNA becomes 2-fold more after each PCR cycle (when 100% PCR efficiency) so that the expression difference is $2^3 = 8$ -fold.

Experimental had 8-fold less *Tar* DNA compared to that of Cont, because high Ct value means less DNA at the starting point.

$$\Delta \text{Ct}^{Ref} = (2^{\text{ExpCt}^{Ref}} / 2^{\text{ContCt}^{Ref}})$$

= $(2^{18} / 2^{19}) = (2^{18-19}) = 2^{-1}$

In the same manner the expression difference is $2^{-1} = 1/2$ -fold, meaning Exp had 1/2 times less (meaning 2-fold more) *Ref* DNA compared to that of Cont at the starting point.

This is also assuming that *Ref* has 100% PCR efficiency.

$$\Delta\Delta Ct = (\Delta Ct^{Tar}/\Delta Ct^{Ref}) = (2^3/2^{-1}) = 2^{3-(-1)} = 2^4 = 16$$

This is compensating the amount of *Tar* using the amount of *Ref* gene.

Real expression difference is 16-fold.

This means the amount of Tar in Exp was 16-fold less than that in Cont so that if you set the amount of Tar in Cont as 1.0, the amount of Tar in Exp is 0.0625 (= 1/16).

Attention!!:

This calculation is applicable only when the PCR efficiency of both *Tar* and *Ref* is 100% (DNA product becomes 2-fold after each PCR cycle).

If the PCR efficiency is less than 100%, you have to change the calculations.

Hypothetical results:

The efficiency was 95% for *Tar* and 93% for *Ref*, meaning that DNA becomes 1.95-fold more for *Tar* and 1.93-fold more for *Ref* after each PCR cycle.

The calculations change as follows:

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\Delta \text{Ct}^{Tar} = (1.95^{\text{ExpCt}^{Tar}} / 1.95^{\text{ContCt}^{Tar}}) = (1.95^{25} / 1.95^{22})
= (1.95^{25-22}) = 1.95^{3}
\Delta \text{Ct}^{Ref} = (1.93^{\text{ExpCt}^{Ref}} / 1.93^{\text{ContCt}^{Ref}}) = (1.93^{18} / 1.93^{19})
= (1.93^{18-19}) = 1.93^{-1}
\Delta \Delta \text{Ct} = (\Delta \text{Ct}^{Tar} / \Delta \text{Ct}^{Ref}) = (1.95^{3} / 1.93^{-1})
= (7.41 / 0.52) = 14.31
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This means that the amount of *Tar* in Exp was 14.31-fold less than that in Cont so that if you set the amount of *Tar* in Cont as 1.0, the amount of *Tar* in Exp is approximately 0.070 (= 1/14.31)

Important rules:

You can use the $\Delta\Delta$ Ct method only when the PCR efficiency of *Tar* and *Ref* is steady, high and almost equal.

How to evaluate the PCR efficiency?

- When you make calibration curves for *Tar* and *Ref*, you will have values of the steadiness (R²), the amplification efficiency and the slope for each calibration curve (refer to the next page).
- If the efficiency is 90% or higher and R^2 is 0.95 or higher (0.99 or higher is preferred) for both Tar and Ref and at the same time the difference of slope values between Tar and Ref is less than 0.1 (means 0.999 is OK but 0.100 is not OK), the Experimentalal condition is good enough to use the $\Delta\Delta$ Ct method, otherwise you have to use the calibration curve method.

